

In Vitro EpiDerm™ Skin Corrosion Test (EPI-200-SCT)

For use with MatTek Corporation's Reconstructed Human Epidermal Model EpiDerm™

Note: MatTek adaptation of this SOP MK-24-007-0024 was based on Zebet's SOP May 7th 2006 (drafted by Manfred Liebsch and Dieter Traue of ZEBET at the BfR and approved by Helena Kandarova)

This SOP is based on the SOP of the validated and regulatory accepted EpiDerm Skin Corrosion Test (SCT). The current SOP updates version from 24th October 1997 and takes into account some "lessons learnt" after the validation of the EpiDerm, EPISKIN models, mainly the correction of direct MTT reduction by the test chemical.

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1. Rationale and Background

Skin corrosion refers to the production of irreversible tissue damage in the skin following the application of a test material, as defined by the Globally Harmonized System (GHS) for Classification and Labeling of Chemical Substances and Mixtures (1)

The potential for chemical induced skin corrosion is an important consideration in establishing procedures for the safe handling, packing and transport of chemicals. Various systems for classification of corrosive potential are included in international regulatory requirements.

The present test is based on the experience that corrosive chemicals are cytotoxic after a short term exposure to the stratum corneum of the epidermis, if cytotoxicity is immediately determined after chemical exposure. It is designed to predict and classify skin corrosivity potential of a chemical by using a three dimensional human epidermis model.

In the year 1998 the EPISKIN and TER *in vitro* corrosivity test were successfully validated and met the acceptance criteria previously defined by the Management Team of the ECVAM International Validation Study (2). Because EPISKIN was not available after the Study, a Catch up validation study was performed with the epidermis model EpiDerm (3).

In 2002 National Co-ordinators of OECD Test Guideline Programme (WNT) endorsed New Draft Test Guidelines TG430 (TER) and TG431 (Human Skin Model) for *In Vitro* Skin Corrosion Testing which were finally adopted in 2004. In Guideline TG 431 (paragraphs 9 – 11) general functional and performance criteria were defined if other (or new) skin or epidermis models are used in the context of this guideline (4).

2. Specific Purpose of the Method

This test is able to reliably discriminate chemicals that are corrosive to skin from non-corrosive chemicals, and can therefore be used for the classification of skin corrosion hazard according to the GHS System adopted by the OECD (1). The Test is not designed to distinguish between sub-categories of corrosivity. However, 70% of the chemicals that are corrosive *in vivo* after 3 minutes exposure already, are also corrosive in the skin model after 3 minutes exposure (3). The test is not designed to predict skin irritation potential.

3. Basic Procedure

EpiDerm tissues are conditioned by pre-incubation (1 hour or overnight) for release of transport stress related compounds and debris. After pre-incubation tissues are transferred to fresh Maintenance Medium and topically exposed with the test chemicals for 3 min and 1 hr, respectively. Two (alternatively three) tissues each are used per treatment, negative control (NC) and positive control (PC). After exposure tissues are rinsed and blotted and assay medium is replaced by MTT-medium. After 3 hr incubation, tissues are washed with PBS, blotted, and the blue formazan salt is extracted with MatTek extractant solution. The optical density (OD) of the formazan extract is determined spectrophotometrically at 540 -570 nm, and cell viability is calculated for each tissue as % of the mean of the negative control tissues. Skin corrosivity potential of the test materials is classified according to the remaining cell viability obtained after 3 minutes or 1 hour exposure with the test chemical.

4. Limitations of the Method

One limitation is possible interference of the test substance with the endpoint MTT: A test substance may directly reduce MTT, thus mimicking dehydrogenase activity of the cellular mitochondria. This property of the test substance is only a problem, if at the time of the MTT test (after the chemical has been rinsed off) there is still sufficient amounts of the test substance present on (or in) the tissues. In this case the (true) metabolic MTT reduction and the (false) direct MTT reduction can be differentiated and quantified by a procedure described in section 6.2.

The method is not designed to be compatible with highly volatile test substances. However, possible toxic interference across plate wells can be avoided by sealing the wells with an adhesive cover sheet, or testing volatile chemicals on separate plates.

5. Materials

5.1 Materials Required for the Experiments

Sterile, blunt-edged forceps	For transferring tissues from agarose
500 mL wash bottle	For rinsing tissue after test material exposure
200 ml beaker	For collecting PBS washes
Sterile disposable pipettes, pipette tips and pipetters	For diluting, adding, and removing media and test materials. For topically applying test materials to tissues
37°C incubator 5% CO ₂	For incubating tissues prior to and during assays
Vacuum source/trap (optional)	For aspirating solutions
Laminar flow hood (optional)	For transferring tissues under sterile conditions
Cotton swabs	For drying the tissue surface
Mortar and Pestle	For grinding solids
Adjustable Pipette 1 mL	For pipetting assay medium under inserts (1 mL)
Pipette 300 µL	For pipetting MTT medium into 24-well plates
Pipette 2 mL	For pipetting MTT extraction solution into 24-well plate
Pipette 200 µL	For pipetting extracted formazan from 24-well plate into 96 well plate to be used in a plate photometer
Pipette 50 µL	For application of liquid test materials
Positive displacement pipette 50 µL	For application of semi-solid test materials
Sharp spoon (NaCl weight: 25 ± 1 mg)	For application of solids Aesculap, Purchase No.: FK 623 MEDKA KG Arzt- und Krankenhausbedarf Bismarrckstr. 101 D –10625 Berlin
bulb headed Pasteur pipette	To aid leveling the spoon and spreading the chemical
Laboratory balance	For pipette verification and checking spoonful weight
96-well plate photometer 570 or 540 nm	For reading OD
Shaker for microtiter / MILLICELL® plates	For extraction of formazan
Stop-watches	To be used during application of test materials
Potassium Hydroxyde, 8 N (Sigma P4494)	To be used as positive control with each kit
Dulbecco's Phosphate Buffered Saline (TC-PBS)	Use for rinsing tissues
MTT (MTT-100-CON)	For preparing MTT-medium

Protocol: In Vitro EpiDerm™ Skin Corrosion Test (EPI-200-SCT)

HCl	For pH adjustment of PBS (if applicable)
NaOH	For pH adjustment of PBS (if applicable)
Nylon Mesh 200 µm (EPI-MESH)	Use as a spreading aid for liquid test materials, provided a pre-test shows the compatibility of test material and nylon mesh and only for chemicals which are not mechanically spread.
H ₂ O, pure (distilled or aqua-pur) Extractant solution (MTT-100-EXT)	To be used as negative control with each kit For MTT extraction

5.2 EpiDerm™ Skin Corrosion Test Kit Components

EPI-200-SCT kits are shipped from Boston usually overnight on Monday (an alternative shipping procedure – over weekend - is possible). Upon receipt of the EpiDerm tissues, place the sealed 24 well plates and the assay medium into the refrigerator (2-8°C). Place the MTT concentrate containing vial in the freezer (-20°C) and the MTT diluent in the refrigerator (2-8°C).

Record lot numbers of all kit components in the Methods Documentation Sheet (MDS) (see ANNEX A).

EPI-200-SCT Kit Components

Amount	Item	Contains/Used for:
1	Sealed 24-well plate (EPI-200-SCT)	Contains 24 tissues on agarose
2	24-well plates (sterile)	For MTT viability assay
4	6-well plates (sterile)	For corrosivity assay
1 bottle, 100 ml	Assay Medium (EPI-100-ASY)	For corrosivity assay
2 bottle, 100 ml	PBS Rinse Solution (TC-PBS)	For rinsing tissues
24 pieces	Nylon Mesh circles 8 mm diameter, 200 µm pore (EPI-MESH)	For spreading test chemicals
1	MK-24-007-0024	Skin corrosivity test (SCT) protocol
1 vial, 10 ml	1% Triton X-100 Solution (TC-TRI)	Skin irritant reference chemical Do not use in present method

MTT-100 Assay Kit Components

1 vial, 2 ml	MTT concentrate (MTT-100-CON)	Frozen MTT concentrate
1 vial, 8 ml	MTT diluent (MTT-100-DIL)	For diluting MTT concentrate prior to use in the MTT assay
1 bottle, 60 ml	Extractant Solution (MTT-100-EXT)	For extraction of formazan

Expiration and Kit Storage

Part #	Description	Conditions	Shelf life*
EPI-200-SCT*	EpiDerm cultures	refrigerator (2-8°C)	96 hours
EPI-100-ASY	Assay medium	refrigerator (2-8°C)	7 days
MTT-100-DIL**	MTT diluent	refrigerator (2-8°C)	2 months
MTT-100-CON**	MTT concentrate	freezer (-20 ± 5°C)	2 months
*Refers to storage time @ 2-8°C in unopened package. ** MTT-100 kits must be ordered separately.			

Note : Examine all kit components for integrity. If there is a concern call MatTek immediately. Contact persons:
 Mitch Klausner, ☎ +1-508-881-6771, Email: mklausner@mattek.com
 Paul H. Kearney, ☎ +1-508-881-6771, Email: phkearney@mattek.com
 Helena Kandarova, ☎ +421-2-3260-7401, Email: hkandarova@mattek.com

6. Methods

6.1 Tissue and Medium Storage

For the EpiDerm reconstructed epidermal model and Medium refer to the relevant Technical Data and Safety Sheet, located in the plastic file inside the shipping box.

If the test is not performed on the day of receipt, store the EpiDerm™ tissues in the refrigerator at 4°C until next day. If you plan to determine any additional endpoints to MTT viability measurements, place the tissues immediately upon arrival into the EpiDerm Maintenance Medium and pre-incubate 1 hour (37°C, 5% CO₂, humidified atmosphere). Afterwards replace the medium and continue with overnight pre-incubation.

Store Maintenance Medium at 4°C in the dark. The shelf life is limited (see Technical Data file in the shipping box). Use EpiDerm Maintenance Medium at room temperature, do not pre-heat. Record lot numbers of all components in the Methods Documentation Sheet (see ANNEX A)

6.2 Test for Interference of Chemicals with MTT Endpoint and Correction Procedures

As specified in Section 4, a test substance may interfere with the MTT endpoint if: a) it is colored and/or b) able to directly reduce MTT (for possible combination of interactions, see **Annex D**). The MTT assay is affected only if the test material is present in the tissues when the MTT viability test is performed.

Some non-colored test materials may change into colored materials in wet or aqueous conditions and thus stain tissues during the 60 min exposure. Therefore, before exposure, a functional check for this possibility should be performed (**Step 1**).

Step 1

Add **50 µl** (liquid) or **25 mg** (solid - using a sharp spoon as per Section 7.2) of the test substance into 0.3 ml of deionized water. Perform the test in a transparent, preferably glass test-tube since plastic test tubes may react with the test articles during the incubation time. Incubate the mixture in the incubator (37±1°C, 5±1 % CO₂, 95% RH) for 60 min. At the end of the exposure time, shake the mixture and evaluate the presence and intensity of the staining (if any). If the solution changes color significantly, the test substance is presumed to have the potential to stain the tissue. A functional check on viable tissues should be performed (**Step 2**).

Step 2

To check the tissue-binding of a **colored test article** (or a chemical that changes into a colored substance), expose one viable tissue to **50 µl** of liquid or **25 mg** of solid test substance. In parallel, expose a tissue to DPBS (negative control). Follow all procedures as described in this SOP Section 6.7 except incubate the tissue for 3-h incubation in culture media without MTT ($37\pm 1^\circ\text{C}$, $5\pm 1\%$ CO_2 , 95% RH) instead of incubating in media containing MTT. After the 3 hour incubation, rinse the tissues and extract the tissues using 2.0 ml of isopropanol and measure the optical density (OD) at 570 nm.

Note: If the colored test substance does not completely rinse off, pipette 1.0 ml of the extracting agent into each well so that the MTT is extracted through the bottom of the tissue culture insert. After extraction is complete, remove the insert and add an additional 1.0 ml of extractant to bring the total volume to 2.0 ml.

Data correction procedure – colored substances

If the extract from tissues treated by colored substance (or substance detected in step 1) has an OD between 5% and 30% of the negative control tissue (treated with PBS), the chemical should be further tested on more tissues using the procedure described above. The real MTT OD (unaffected by interference with the colored test materials) is calculated using following formula:

$$\text{OD} = \text{OD colored tissue (MTT assay)} - \text{OD colored tissue (no MTT assay)}$$

Note: If the extract from tissues treated by colored substance (or substance detected in step 1) has an OD <5% of the PBS treated control tissue and the tissue viability (determined in MTT assay) is not close to the classification cut-off (50%), correction of the results is not necessary.

If OD of extract from the tissue treated by colored substance (or substance detected in **Step 1**) is > 30% of the PBS treated control tissue, additional steps and expert judgment must be performed to determine if the test substance must be considered as incompatible with the test.

Step 3

All test materials (including those already evaluated in **Step 1** and **Step 2**) should be further evaluated for their **potential to interfere with MTT assay**. To test if a material directly reduces MTT, add **50 µl** (liquid) or **25 mg** (solid - using sharp spoon **7.2**) of the test substance to 1 ml of the MTT medium and incubate in the incubator ($37\pm 1^\circ\text{C}$, $5\pm 1\%$ CO_2 , 95% RH) for 60 min. Untreated MTT medium is used as control. If the MTT solution turns blue/purple, the test substance reduces MTT and additional functional check (**Step 4**) must be performed.

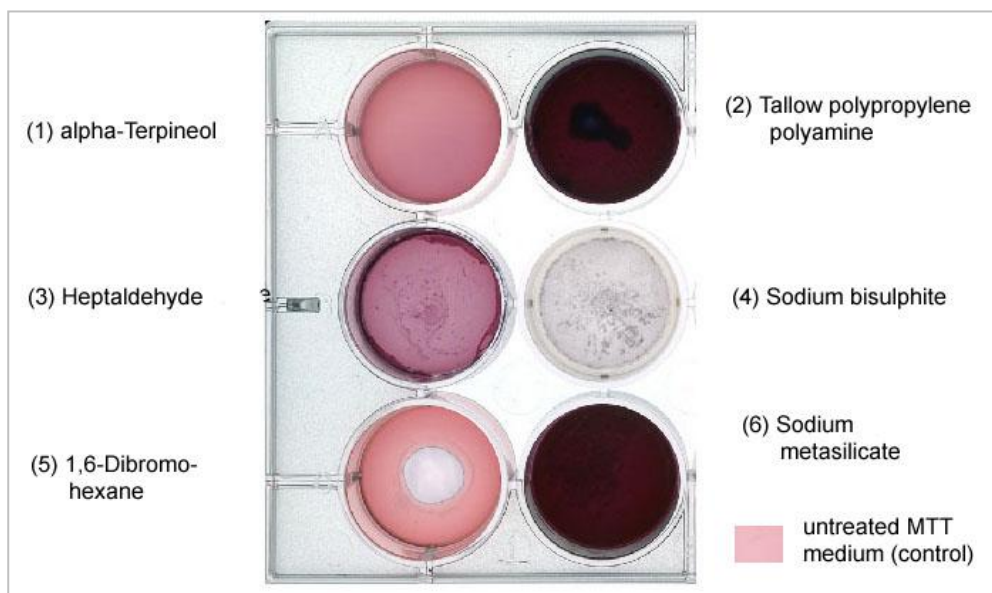


Figure 1: Example of test for direct MTT reduction ability (Step 3). Test substances (2) (3) and (6) have directly reduced MTT. In these cases, Step 4 (below) must be performed.

Step 4:

The procedure employs freeze-killed tissues that possess no metabolic activity but absorb and bind the test substance similar to viable tissues.

- Freeze-killed tissues can be ordered separately from MatTek Corporation (part # EPI-200-FRZN-EA). The frozen tissues may be stored indefinitely in the freezer (-20 ± 5°C).
- Each MTT reducing chemical is applied to two freeze-killed tissues. In addition, two freeze killed tissues are left untreated (Note: The untreated killed controls will show a small amount of MTT reduction due to residual reducing enzymes within the killed tissue). The entire assay protocol is performed on the frozen tissues in parallel to the assay performed with the live EpiDerm tissues. Data are then corrected as follows:

Data correction procedure – MTT reducers

True viability = Viability of treated tissue – Interference from test chemical = OD tvt – OD kt
where OD kt = (mean OD tkt – mean OD ukt)

tvt = treated viable tissue
tkt = treated killed tissue

kt = killed tissues
ukt = untreated killed tissue (NC treated tissue)

If the interference by the test substance is greater than 30% of the negative control value, additional steps must be taken into account or the test substance may be considered incompatible with this test system (expert judgment).

If the interference by the test substance is < 30% of the negative control value, the net OD of the test substance treated killed control may be subtracted from the mean OD of the test substance treated viable tissues to obtain the true amount of MTT reduction that reflects metabolic conversion only.

Note 1: If the colored test material or the MTT reducing chemical is classified as irritant by the SIT (tissue viability <50 %), the correction procedures are not necessary.

Note 2: Frozen tissues (EPI-200-FRZN-EA) can be ordered separately from MatTek Corporation.

6.3 Test for Mesh Compatibility (Liquid Test Substances Only)

Since the surface of the EpiDerm tissues is slightly hydrophilic, spreading of highly lipophilic substances might sometimes present a problem due to the surface tension effects. Therefore, a nylon mesh (see 5.1) can be used as a spreading support if mechanical re-spreading is not efficient. Some chemicals, however, may react with the mesh and therefore the compatibility of test chemical with nylon mesh must be performed. To test if a test chemical interacts with the mesh, place the mesh on a glass slide and apply 50 µL of test substance. After 60 minutes exposure, check using a microscope. If an interaction between test substance and the mesh is noticed (Figure 2), the test substance has to be applied without the mesh.

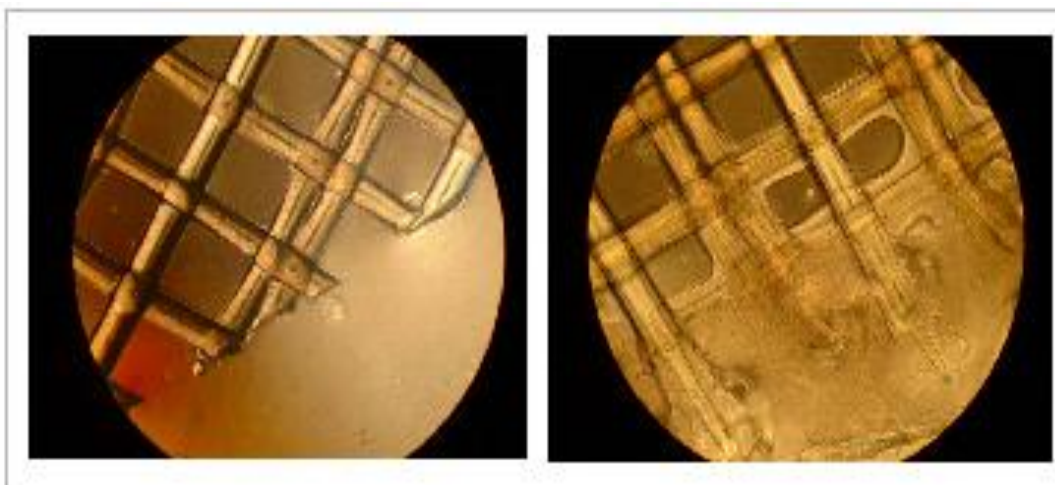


Figure 2: The mesh compatibility test

Note: The mesh (25 pieces) is included in the EPI-200-SCT kit from MatTek.

6.4 Preparations

6.4.1 MTT Medium (Prepare Fresh on Day of Testing!)

Thaw the MTT concentrate (MTT-100-CON) and dilute with the MTT diluent (MTT-100-DIL). Store the remaining MTT solution in the dark at 4°C for later use on the same day (do not store until next day since MTT will degrade with time).

After filtration (using a sterile 0.45µm filter) add 2 mL of the stock-solution to 8 ml Maintenance Medium (final concentration: 1 mg MTT / ml medium). Keep the MTT medium in the dark. Record the preparation in the MDS.

MTT Stock solution (prepared in PBS) can be stored frozen (-20 °C) up to one month.

Safety precaution: MTT is toxic (Risk phrases: R26, R68, R22, R36, R37, R38). Wear protective gloves during manipulation with MTT solution!

Note: MTT is light sensitive. Protect all solutions from light

6.4.2 Dulbeccos PBS

Sterile ready-to-use DPBS should be used. About two liters are sufficient for all rinsing performed with one kit. If PBS is prepared from powder or 10x concentrated PBS is used, prepare according to supplier instructions and adjust to pH 7.0 with either NaOH or HCl. Record the pH adjustment in the MDS

Note: Alternatively, ready-to-use PBS can be used. About one litre is sufficient for all rinsing performed with 24 tissues.

6.4.3 Test Substances

Safety Instruction

- a) For handling of non-coded test substances follow instructions given in the Material Safety Data Sheet.
- b) If coded chemicals are supplied, no (or possibly incomplete) information regarding the safe handling will be provided. Therefore, all test materials must be treated as if they were corrosive and toxic and work must be performed in accordance with chemical safety guidelines (use ventilated cabinet, wear gloves, protect eyes and face).
- c) Store all test substances according to recommendations. Respect special store conditions (special temperature, protection from light, protection from oxidization by nitrogen, etc.)

Liquids: Dispense **50 µL** directly atop the EpiDerm tissue and spread with bulb headed Pasteur pipette. If necessary gently place a nylon mesh (8 mm diameter) on the surface (see Figure 3, picture b).
Record the use of mesh as spreading tool in the MDS.

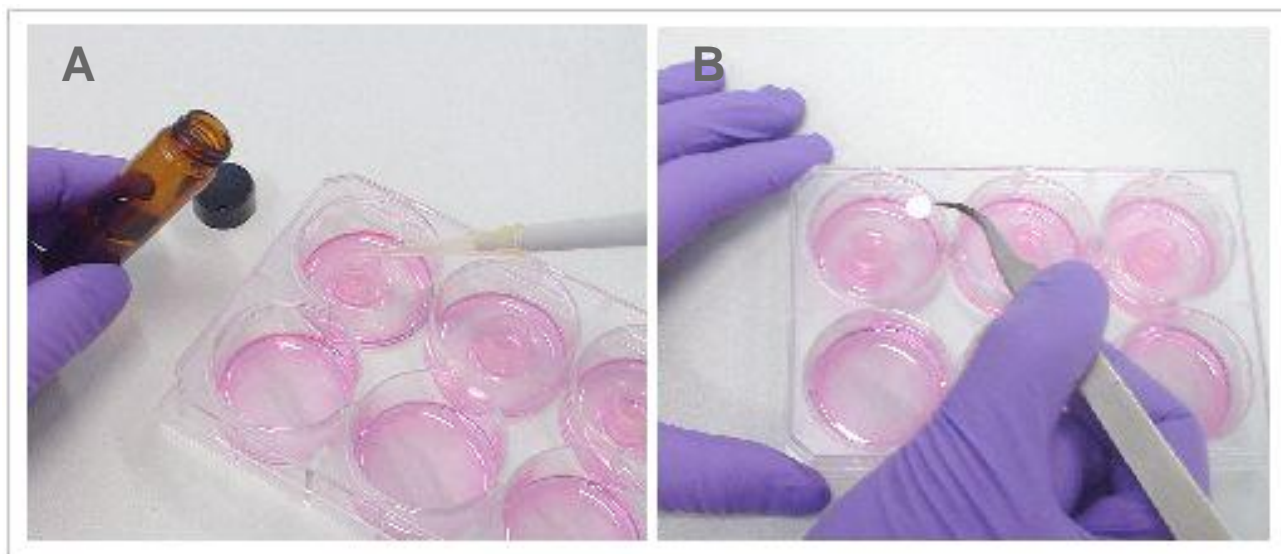


Figure 3: Application of liquids.

Semisolids: Dispense **50 µl** using a positive displacement pipette directly atop the tissue. If necessary spread to match size of tissue. *Record the use of spreading in the MDS.*



Figure 4: Application of semisolids.

Solids: Crush and grind test material with a mortar with pestle wherever this improves the consistency. Fill **25 mg** sharp application spoon* (see **5.1** and **Figure 5**) with fine ground test material. Level the spoon by gently scratching the excess material away with an appropriate aid, avoiding compression ("packing") of the test material. "Packing" can be avoided by using a rod shaped sound instead of a flat spatula. If a bulb headed sound is used the bulb can be used to empty the spoon completely. Add the **25 µl of H₂O** necessary for wetting of the test material to increase tissue surface contact. Increase the volume of H₂O in case of materials where this is not enough for wetting. If necessary spread material to match the size of the tissue. Record in the MDS if grinding was not used and the H₂O volume necessary to wet the chemical.



Figure 5: Application of solids.

Waxes: For test substances with **waxy consistence** the spoon application does not work. In these cases try to form a flat “disc like” piece of about 8 mm diameter and place it atop the tissue, wetted with **15 µl H₂O**. To improve the contact between test substance and tissue weigh down the “disc” with a stainless steel aid like that shown in **Figure 6**.



Figure 6: Stainless steel aid.

**Note: Since the surface of the solid covering the tissues is more important than the weight, the “leveled spoon technique” is an accepted dosing procedure. The spoon used here has been calibrated to equal 25mg of fine grinded NaCl. The weight will be different if other materials are used.*

****Note:** determine in a pre-test a volume of H₂O necessary to wet test chemical.

6.5 Prediction Model

Corrosivity potential of the test materials is predicted from the relative mean tissue viabilities obtained after a **3 min** treatment compared to the negative control tissues concurrently treated with H₂O. A chemical is classified "**corrosive**" in any case, if the relative tissue viability after **3 min** treatment with a test material is decreased below 50%. In addition, those materials classified "**non corrosive**" after **3 min** (viability ≥ 50%) are classified "**corrosive**" if the relative tissue viability after **1 hr** treatment with a test material is decreased below 15%.

Mean tissue viability (expressed as % of negative control)	Prediction
3 min < 50%	corrosive
3 min ≥ 50% and 1 hour: < 15%	corrosive
3 min ≥ 50% and 1 hour: ≥ 15%	non-corrosive

6.6 Assay Quality Controls

6.6.1 Assay Acceptance Criterion 1: Negative Control (NC)

The **absolute OD** of the H₂O treated NC tissues in the MTT-test is an indicator of tissue viability obtained in the testing laboratory after the shipping and storing procedure and under specific conditions of the assay. Tissue viability is meeting the acceptance criterion if the mean OD of the mean of NC is **OD ≥ 0.8**.

6.6.2 Assay Acceptance Criterion 2: Positive Control (PC)

8N KOH (Sigma # P4494) is used as PC and has to be tested once on each testing day. 1 hour exposure of the PC will reveal a mean tissue viability less than **15 %**.

6.6.3 Assay Acceptance Criterion 3: Coefficient of Variation (CV)

Based on the test design (explained in 6.6) the experiments are performed on two or three tissue replicates per exposure time. In the range between 20% and 100% viability the coefficient of variation (CV) is an additional acceptance criterion. **It should not exceed 0.3.**

6.7 Experimental Procedure

Introductory Note:

The experiment can be performed in 2 modification using a) two tissues per timepoint (as in the validation study) or b) three tissues per timepoint. Alternative b) is recommended mainly for laboratories establishing the test. Examples of both test design are given below.

ALTERNATIVE A) 24 tissues (1 EPI-200-SCT kit) are used for testing **4 test chemicals**, negative control and positive control, each of them applied both for **3 min** and **1 hr** to **two tissue replicates** per treatment.

ALTERNATIVE B) 36 tissues (1 EPI-200-SCT and 1 EPI-212-SCT kit) are used for testing **4 test chemicals**, negative control and positive control, each of them applied both for **3 min** and **1 hr** to **three tissue replicates** per treatment..

Day Prior To Testing

- If the EpiDerm tissues are not used in the day of receipt, keep the originally sealed plates until the next day in the refrigerator at 4°C.
- If necessary, prepare sufficient amount of rinsing PBS for the next day according to **6.4.1**

Note: Since the present test is a short term test which makes use of the EpiDerm model over a period of only 5 hours, sterility is not as important as is in other applications of the EpiDerm model. Nevertheless, it is important to keep assay media sterile and to keep risk of contamination at a low level

ALTERNATIVE A) – Example With 4 Chemicals, Two Tissues Per Timepoint

Day of Testing

Tissue Conditioning (Pre-incubation)

- a) Prepare **two** 6-well plates for **four** chemicals and the negative (NC) and positive control (PC) for the **3 min application**. Pipette 0.9 mL Maintenance Medium in each well.
- b) Prepare **two** 6-well plates for **four** chemicals and the negative (NC) and positive control (PC) for the **1 hr application**. Pipette 0.9 ml Maintenance Medium in each well.
- c) Remove the shipped multiwell plate from the plastic bag. Open the 24-well plate under a sterile airflow and remove the sterile gauze. Carefully take out each insert containing the epidermal tissue, rapidly remove any remaining agarose that adheres to the outer sides of the insert by gentle blotting on the sterile filter paper, and immediately place it in a well of the prepared 6-well plate. Act quickly as the epidermal cultures dry out rapidly when not in contact with medium. Make sure that no air bubbles are formed underneath the insert!
- d) Mark the 6-well plates as shown in Figure 7.
- e) Place the 6-well plates containing the tissues into a humidified (37°C, 5% CO₂) incubator for 1 hour.

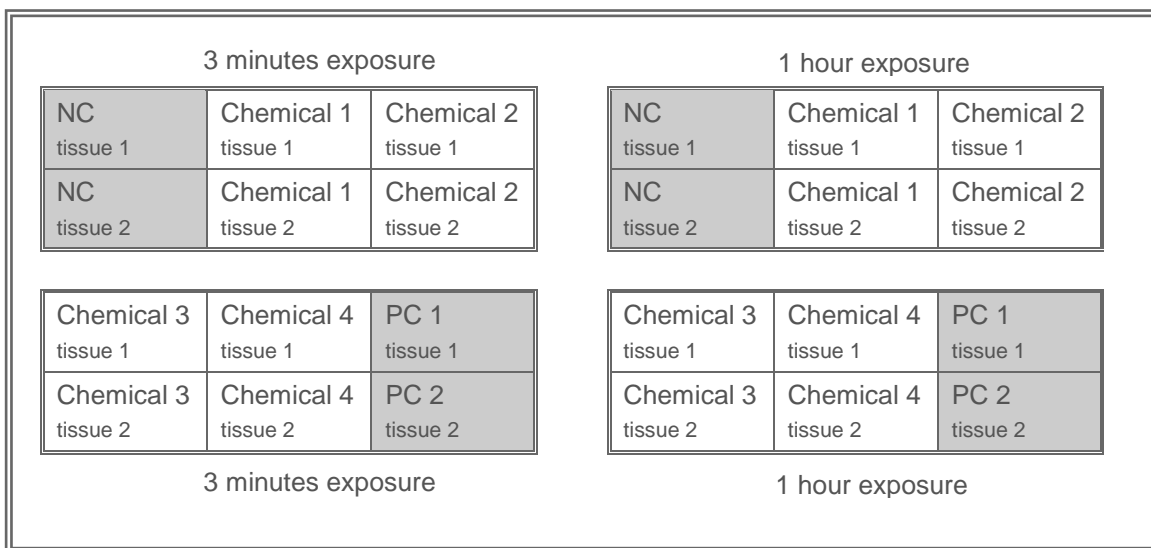


Figure 7: 6-well plate design

Preparations for the Main Test (Make During the 1 Hr Pre-incubation Time)

- a) Prepare the MTT medium according to 6.4.1.
- b) Prepare two 24-well plates to be used as “holding and “MTT plates” one for the 3 min experiment, the other for the 1 hr experiment
- c) Use the plate design shown in Figure 8. Pipette 300 µl of either maintenance medium or MTT medium in each well.
- d) Place the plates in the incubator.

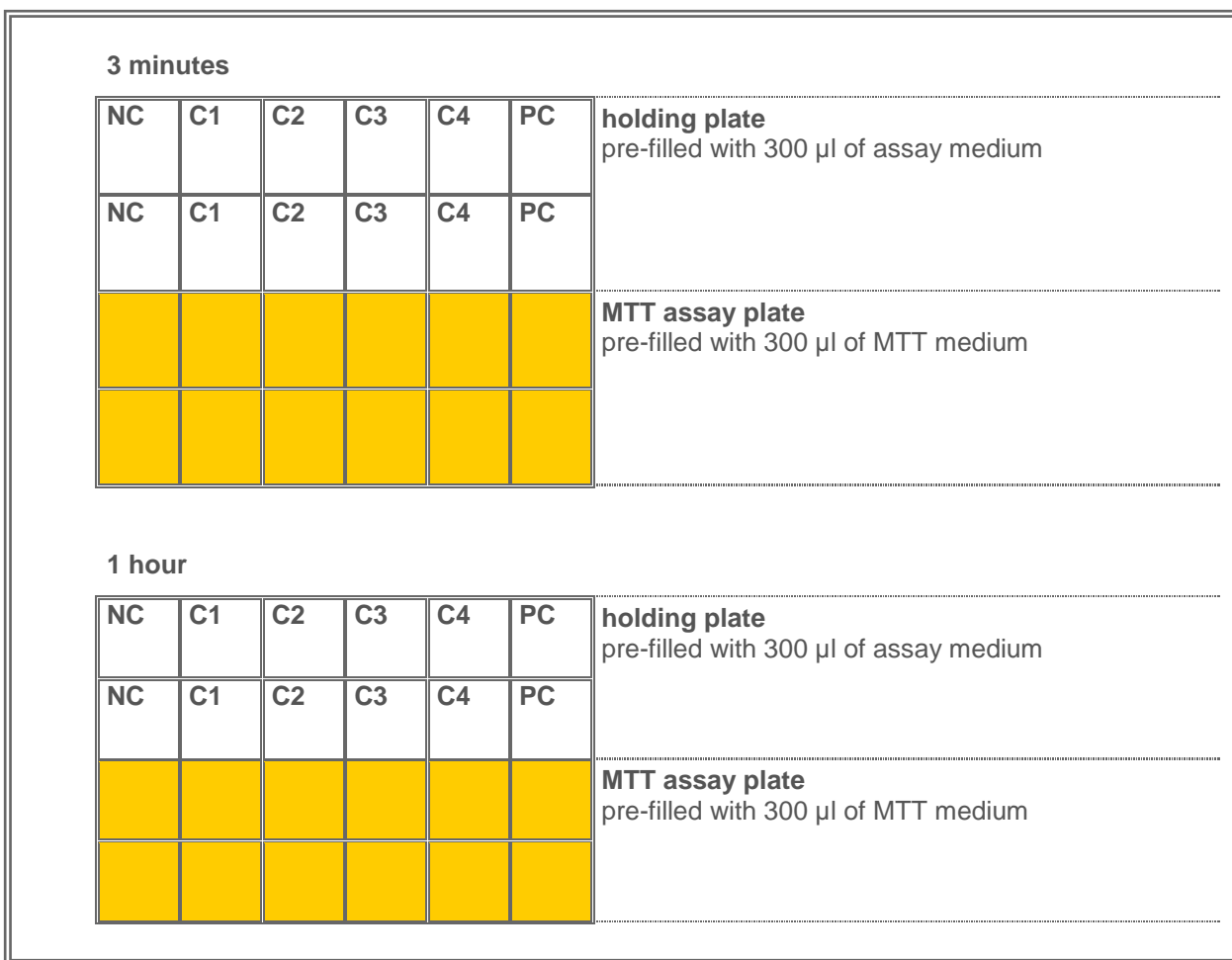


Figure 8: 24-well plate design. "Holding and MTT plate", both for the 1 hr experiment and the 3 min experiment. NC = negative control; C1 – C4 = test chemicals 1-4; PC = positive control.

1-Hour Application

Note: Dosing time interval is dictated by rinsing procedure. If the technician has performed the test already, 45 seconds intervals is sufficient for both application and washing procedures. However, if the test is performed for the first time, 1 minute dosing interval is recommended.

- a) After 1 hour of pre-incubation, transfer each insert to new 6-well plates with fresh medium (0.9 ml per well). Alternatively, aspirate the pre-incubation medium from the 6-well plates and pipet 0.9 ml of fresh medium into each well.
- b) Set the timer to 1 hr and start it. Add 50 µl of H₂O (negative control) into the first insert atop the tissue. After 45 sec repeat the procedure with the second tissue. Proceed with test material 1 - 4 (50 µl: liquids, 25 mg + 25 µl H₂O: solids) and the positive control in the same manner until all tissues are dosed.
- c) Dosing interval scheme for the 3 min experiment:

0.00-0.45 – tissue 1 (NC)	4.30-5.15 – tissue 7 (C3)
0.45-1.30 – tissue 2 (NC)	5.15-6.00 – tissue 8 (C3)
1.30-2.15 – tissue 3 (C1)	6.00-6.45 – tissue 9 (C4)
2.15-3.00 – tissue 4 (C1)	6.45-7.30 – tissue 10 (C4)
3.00-3.45 – tissue 5 (C2)	7.30-8.15 – tissue 11 (PC)
3.45-4.30 – tissue 6 (C2)	8.15-9.00 – tissue 12 (PC)
- d) Place the 6-well plates into the incubator (37°C, 5% CO₂) for the rest of the exposure time until 1 hour exposure is reached for first tissue dosed. Record start time in the MDS.
- e) After the 1 hour period of test material exposure is complete, with forceps remove the first insert from the 6-well plate. As shown in Figure 9, using a wash bottle gently rinse the tissue with PBS (= fill and empty insert 20 times in a constant soft stream of PBS) to remove any residual test material. Remove excess PBS by gently shaking the insert and blot bottom on blotting paper. Place the insert in the prepared holding plate. Proceed with test materials 1 - 4 and the positive control in the same manner until all tissues are rinsed. Rinse all tissues in an interval of 45 sec.



Figure 9: Rinsing: fill and empty 20 x

- f) Once all tissues have been rinsed and are in the holding plate, dry the surface with cotton swab, remove inserts from the holding plate, blot bottom and transfer into the 24-well plate, prepared for the MTT assay.
- g) Place plate in the incubator, record start time of MTT incubation in the MDS and incubate for 3 hours (37°C, 5% CO₂).

3 Min Application

Note: Dosing time interval is dictated by rinsing procedure. If the technician performs test already routinely, 45 seconds interval is sufficient for both application and washing procedures. However, if the test is performed for the first time, 1 minute dosing interval is recommended.

- a) Start the timer for 3 min. Add 50 µl H₂O (negative control) into the first insert atop the tissue. After 45 seconds repeat the procedure with the second tissue. Following 45 seconds intervals enable to dose 4 tissues. Afterwards, washing of the tissue 1 have to start.
- b) After the 3 min period of exposure for the first four tissues is complete, start the timer for 3 min and with forceps remove the first insert from the 6-well plate. As shown in Figure 9, using a wash bottle gently rinse the tissue with PBS (= fill and empty insert 20 times in a constant soft stream of PBS) to remove any residual test material. Remove excess PBS by gently shaking the insert and blot bottom on blotting paper. Place insert in the prepared holding plate. After 45 seconds repeat the procedure with the second insert, after 1:30 minutes with the third tissue...etc.

Dosing interval scheme for the 3 min experiment:

SET 1 = NC and chemical 1 (6 minutes)

Dosing:

- 0.00-0.45 – tissue 1 (NC)
- 0.45-1.30 – tissue 2 (NC)
- 1.30-2.15 – tissue 3 (C1)
- 2.15-3.00 – tissue 4 (C1)

Rinsing

- 3.00-3.45 – tissue 1 (NC)
- 3.45-4.30 – tissue 2 (NC)
- 4.30-5.15 – tissue 3 (C1)
- 5.15-6.00 – tissue 4 (C1)

SET 2 = chemical 2,3 (6 minutes)

SET 3 = chemical 4, PC (6 minutes)

- c) Proceed with all test materials (50 µl: liquids, 25 mg + 25 µl H₂O: solid) and the positive control in the same manner until all tissues are dosed and rinsed.
- d) Once all tissues have been rinsed and are in the holding plate, carefully dry the surface of the tissue with a cotton swab. Afterwards remove inserts from the holding plate, blot bottom and transfer into the 24-well plate, prepared for the MTT assay. Place plates in the incubator, record start time of MTT incubation in the MDS and incubate for 3 hours (37°C, 5% CO₂).

ALTERNATIVE B) – Example With 4 Chemicals, Three Tissues Per Timepoint

Day of Testing

Tissue Conditioning (Pre-Incubation)

- a) Prepare three 6-well plates for four chemicals and the negative (NC) and positive control (PC) for the 3 min application. Pipette 0.9 ml Maintenance Medium in each well.
- b) Prepare three 6-well plates for four chemicals and the negative (NC) and positive control (PC) for the 1 hr application. Pipette 0.9 ml Maintenance Medium in each well.
- c) Remove the shipped multiwell plate from the plastic bag. Open the 24-well plate under a sterile airflow and remove the sterile gauze. Carefully take out each insert containing the epidermal tissue, rapidly remove any remaining agarose that adheres to the outer sides of the insert by gentle blotting on the sterile filter paper, and immediately place it in a well of the prepared 6-well plate. Act quickly as the epidermal cultures dry out rapidly when not in contact with medium. Make sure that no air bubbles are formed underneath the insert!
- d) Mark the 6-well plates as shown in Figure 10.
- e) Place the 6-well plates containing the tissues into a humidified (37°C, 5% CO₂) incubator for 1 hour.

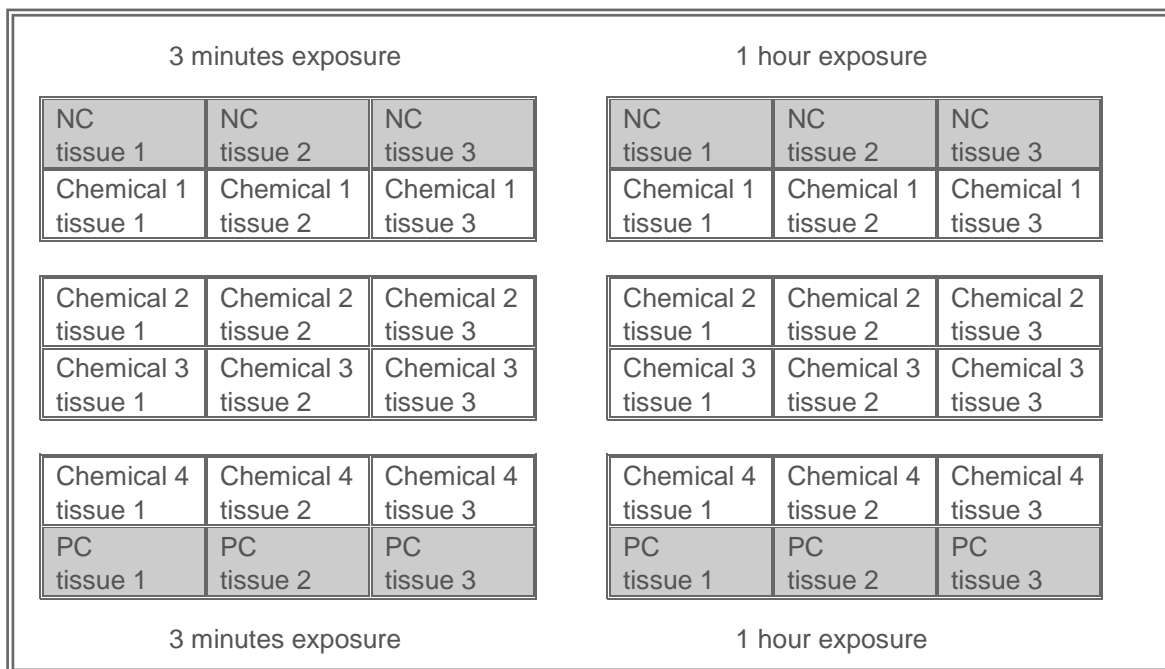


Figure 10: 6-well plate design

Preparations for the main test (make during the 1 hr pre-incubation time)

- a) Prepare MTT medium according to 6.4.1.
- b) Prepare two 24-well plates to be used as “holding plates.” One for the 3 min experiment, the other for the 1 hr experiment. Pipette 300 µl of maintenance medium into each well of the holding plates. In addition, prepare two 24-well plates for the MTT assay. Pipette 300 µl of MTT medium in each well of the MTT assay plate.
- c) Use the plate design shown in Figure 11.
- d) Place the plates in the incubator.

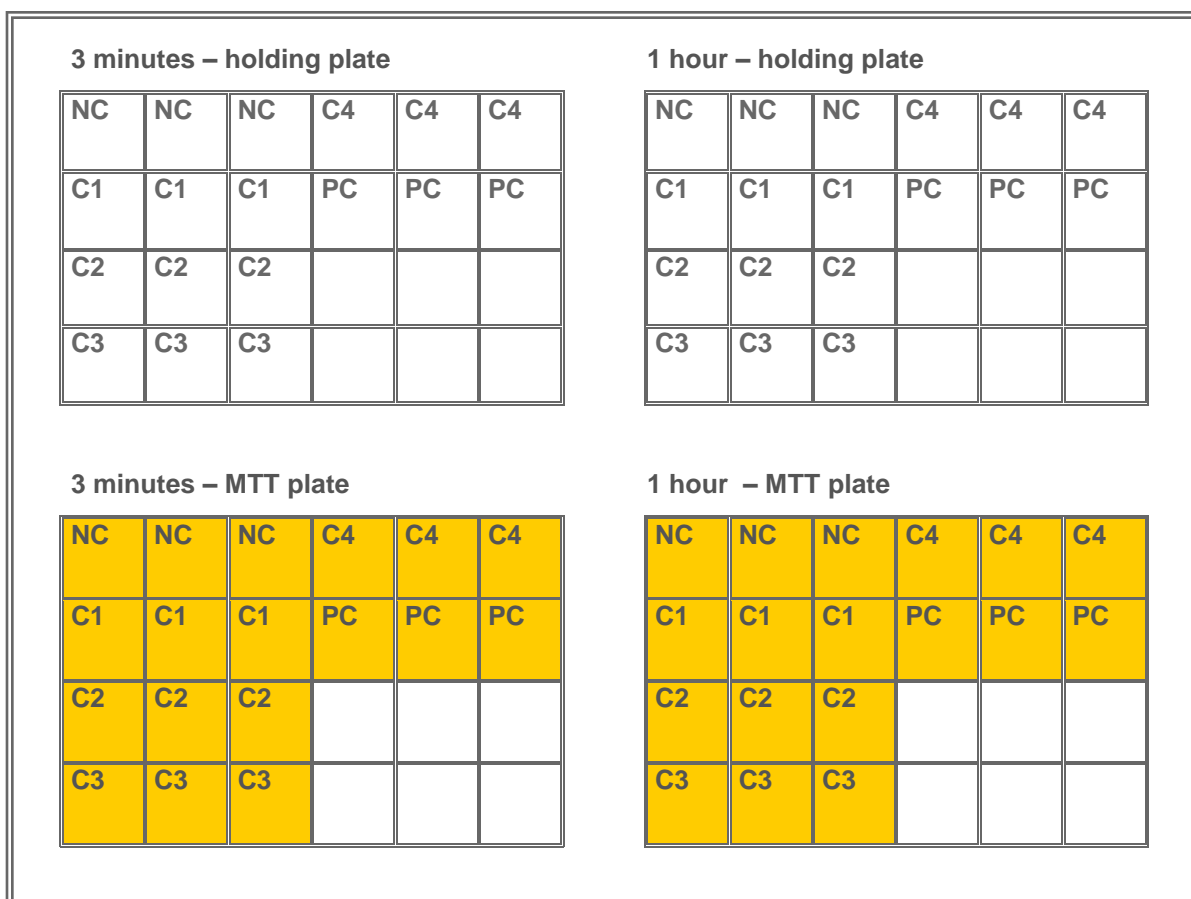


Figure 11: 24-well plate design. "Holding and MTT plates", both for the 3 min and 1 hour experiment. NC = negative control; C1 – C4 = test chemicals 1- 4; PC = positive control

1-hour application

Note: Dosing time interval is dictated by rinsing procedure. In this test design, a 1 minute dosing interval is recommended.

- a) After 1 hour of pre-incubation, transfer each insert to new 6-well plates with fresh medium (0.9 ml per well). Alternatively, aspirate the pre-incubation medium from the 6-well plates and pipet 0.9 ml of fresh medium into each well.
- b) Set a timer to 1 hr and start it. Add 50 µl of H₂O (negative control) into the first insert atop the tissue. After 60 sec repeat the procedure with the second tissue. Proceed with test material 1 - 4 (50 µl: liquids, 25 mg + 25 µl H₂O: solids) and the positive control in the same manner until all tissues are dosed.

Dosing interval scheme for the 1 hour experiment, alternative b):

0.00-1.00 – tissue 1 (NC)	9.00-10.00 – tissue 10(C3)
1.00-2.00 – tissue 2 (NC)	10.00-11.00 – tissue 11 (C3)
2.00-3.00 – tissue 3 (NC)	11.00-12.00 – tissue 12 (C3)
3.00-4.00 – tissue 4 (C1)	12.00-13.00 – tissue 13 (C4)
4.00-5.00 – tissue 5 (C1)	13.00-14.00 – tissue 14 (C4)
5.00-6.00 – tissue 6 (C1)	14.00-15.00 – tissue 15 (C4)
6.00-7.00 – tissue 7 (C2)	15.00-16.00 – tissue 16 (PC)
7.00-8.00 – tissue 8 (C2)	16.00-17.00 – tissue 17 (PC)
8.00-9.00 – tissue 9 (C2)	17.00-18.00 – tissue 18 (PC)

- c) Place the 6-well plates into the incubator (37°C, 5% CO₂) for the remaining exposure time until 1 hour exposure is reached for first tissue dosed. Record start time in the MDS.
- d) After the 1 hour period of test material exposure is complete, use forceps to remove the first insert from the 6-well plate. As shown in Figure 9, using a wash bottle gently rinse the tissue with PBS (= fill and empty insert 20 times in a constant soft stream of PBS) to remove any residual test material. Remove excess PBS by gently shaking the insert and blot the bottom on blotting paper. Place the insert in the prepared holding plate. Proceed with test materials 1 - 4 and the positive control in the same manner until all tissues are rinsed. Rinse all tissues in an interval of 60 sec.
- e) Once all the tissues have been rinsed and are in the holding plate, dry the tissue surface with a cotton swab. Remove inserts from the holding plate, blot bottom and transfer into the 24-well plate, prepared for the MTT assay.
- f) Place the plate in the incubator, record start time of MTT incubation in the MDS and incubate for 3 hours (37°C, 5% CO₂).

3 Min Application

Note: Dosing time interval is dictated by rinsing procedure. In this test design, 1 minute dosing interval is recommended.

- a) Start the timer for 3 min. Add 50 µl of H₂O (negative control) into the first insert atop the tissue. After 60 seconds repeat the procedure with the second tissue. Following 60 seconds interval enable to dose 3 tissues (= 1 test substance).
- b) After the 3 min period of exposure for the first three tissues is complete, remove the first insert from the 6-well plate. As shown in Figure 10, using a wash bottle gently rinse the tissue with PBS (= fill and empty insert 20 times in a constant soft stream of PBS) to remove any residual test material. Remove excess PBS by gently shaking the insert and blot the bottom on blotting paper. Place the insert into the prepared holding plate. After 60 seconds repeat the procedure with the second insert....etc.

Dosing interval scheme for the 3 min experiment:

NC (6 minutes)

Dosing:

0.00-1.00 – tissue 1 (NC)

1.00-2.00 – tissue 2 (NC)

2.00-3.00 – tissue 3 (NC)

Rinsing

3.00-4.00 – tissue 1 (NC)

4.00-5.00 – tissue 2 (NC)

5.00-6.00 – tissue 3 (NC)

chemical 1 = 6 minutes

chemical 2 = 6 minutes

chemical 3 = 6 minutes

chemical 4 = 6 minutes

PC = 6 min

Total duration of the test : 36 min

- c) Proceed with all test materials (50 µl: liquids, 25 mg + 25 µl H₂O: solid) and the positive control in the same manner until all tissues are dosed and rinsed.
- d) Once all the tissues have been rinsed and are in the holding plate carefully dry the tissue surface with a cotton swab. Afterwards remove inserts from the holding plate, blot the bottom and transfer into the 24-well plate, prepared for the MTT assay. Place plate in the incubator, and incubate for 3 hours (37°C, 5% CO₂). Record the start time of MTT incubation on the MDS.

MTT Test and Reading – Both test designs

- a) After the 3 hour MTT incubation period is complete, gently aspirate MTT from all wells (e.g. using a suction pump), refill wells with PBS and aspirate. Repeat the rinsing twice and make sure that tissues are dry after the last aspiration. Transfer inserts to new 24 well plates.
- b) Immerse the inserts by gently pipetting 2 mL extractant solution (MTT-100-EXT) into each insert. The level will rise above the upper edge of the insert, thus completely covering the tissue from both sides.
- c) Seal the 24 well plate (e.g. with a zip bag) to inhibit the extractant solution evaporation. Record the start time of extraction in the MDS. Extract either over night without shaking at room temperature or, alternatively, 2 hours with shaking (~120 rpm) at room temperature.
- d) After the extraction period is complete, pierce the inserts with an injection needle and allow the extract to run into the well from which the insert was taken. Afterwards the insert can be discarded. Place the 24-well plates on a shaker for 15 minutes until solution is homogeneous in colour.
- e) Per each tissue transfer 200 µl of the blue formazan solution into a 96-well flat bottom microtiter plate, both from the 3 min exposure and from the 1 hr exposure. For the 96-well plate, use the exact plate design given in Figure 12, as this configuration is used in the data spreadsheets. Read the OD in a spectrophotometer at 570 nm, without reference filter. Alternatively, ODs can be read at 540 nm.

Note: Readings are performed *without* a reference filter, since the "classical" reference filter often used in the MTT test (630 nm) is still within the absorption curve of formazan. Since filters may have a ± tolerance in some cases the reference filter reduces the dynamics of the signal (OD) up to 40%.

Fixed 96 well-plate design for OD readings

	1	2	3	4	5	6	7	8	9	10	11	12	
A	NC	NC	C1	C1	C2	C2	C3	C3	C4	C4	PC	PC	3 min
B	NC	NC	C1	C1	C2	C2	C3	C3	C4	C4	PC	PC	
C	NC	NC	C1	C1	C2	C2	C3	C3	C4	C4	PC	PC	
D	NC	NC	C1	C1	C2	C2	C3	C3	C4	C4	PC	PC	1 hour
E	NC	NC	C1	C1	C2	C2	C3	C3	C4	C4	PC	PC	
F	NC	NC	C1	C1	C2	C2	C3	C3	C4	C4	PC	PC	
G													
H													
	T1	T2	T1	T2	T1	T2	T1	T2	T1	T2	T1	T2	

Figure 12a. Alternative A): Transfer 3 aliquots (200 µl) from each tissue into the 96-well plates using the plate design above. Use one 96-well plate for both time points.

	1	2	3	4	5	6	7	8	9	10	11	12	
A	NC	C1	C2	C3	C4	PC	NC	C1	C2	C3	C4	PC	T1
B	NC	C1	C2	C3	C4	PC	NC	C1	C2	C3	C4	PC	
C	NC	C1	C2	C3	C4	PC	NC	C1	C2	C3	C4	PC	T2
D	NC	C1	C2	C3	C4	PC	NC	C1	C2	C3	C4	PC	
E	NC	C1	C2	C3	C4	PC	NC	C1	C2	C3	C4	PC	T3
F	NC	C1	C2	C3	C4	PC	NC	C1	C2	C3	C4	PC	
G													
H													
	3 min	3 min	3 min	3 min	3 min	3 min	1 hour	1 hour	1 hour	1 hour	1 hour	1 hour	

Figure 12b. Alternative B): Transfer 2 aliquots (200 µl) from each tissue into the 96-well plates using the plate design above. Use one 96-well plate for both time points.

6.8 Documentation

6.8.1 Method Documentation Sheet, MDS (see ANNEX)

The MDS allows to check the correct set up, calibration and function of the equipment as well as correct weights, applications etc. The MDS is designed as a paper document "in the spirit of GLP". Per each test, make a hardcopy of the MDS, fill in and sign the requested information, starting the day prior to testing and ending after the test has been conducted.

Note (1): If several tests are performed per week, pipette verification (weighing H₂O on a balance) is only necessary once at the beginning of each week. Nevertheless, if adjustable pipettes are used the correct adjustment shall be checked and recorded in the MDS before each test.

Note (2): If solids cannot be sufficiently ground to a fine powder, it is recommended to check the weight of the levelled application spoon and record this weight in the MDS.

6.8.2 MS EXCEL Data Spreadsheets

The MS EXCEL spreadsheets can be obtained from ZEBET or MatTek Corporation, USA. Data of optical densities (ODs) generated by the microplate reader **and already corrected by subtraction of the blanks mean from all OD values** are copied from the Reader software or manually inputted to the Windows Clipboard and then pasted into the first map (Import) of the EXCEL spreadsheet in the 96-well format given above in Figure 12.

The spreadsheet consists of three sheets, named Import, Results and Remarks. The first sheet (**Import**) is used for pasting or manually transposing the OD values. In addition, fill in required information on test substances, tissue lots, etc. in the first sheet (*Import*). The second sheet (**Results**) makes calculations and provides a column graph of the results. Entries on test materials made in the first sheet will be copied from there automatically to the correct positions in the *Results* sheet. Any special comment related to the experiment or measurement can be recorded in third sheet named **Remarks**.

7. REFERENCES

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ANNEX A: Methods Documentation Sheet (MDS)

Assay No:.....
Date:.....
Corresponding XLS data file name:.....
Performed by:.....

Time Protocol

Receipt of EpiDerm tissues:										
Date:.....ID:.....										
Experimental schedule:										
Date:.....ID:.....										
procedure	preincubation (1 hour)		exposure (1hour)		exposure (3 min)		MTT assay (3h)		formazan extraction (2h or overnight)	
SET No.	start	stop	Start	stop	start	stop	start	stop	start	stop
1										
2										
3										
4										
5										
6										
7										
8										
9										
10										
11										
12										

Measurement (date, hour):
Date:.....ID

Devices Verification

Incubator Verification

Incubator #	CO ₂ < 5% ± 0,5 % >	Temperature < 37°C ± 1°C >	Check water in reservoir (✓)

ID/ Date:

Refrigerator Verification

Refrigerator #	Temperature < 5°C ± 2°C >

ID/ Date:

Water Bath Verification

Water bath #	Temperature

ID/ Date:

In case that your devices are controlled by central computer, fill in the following table instead of fields above:

Name of the Device	Device #	Reference

ID/ Date:

Pipette verification (triplicate weightings)

Pipette 3 x H₂O into a small baker on a laboratory scale and record readings in g. Perform pipette verification only once per week and refer to it in all assays of this week. If adjustable pipettes are used, check adjustment daily.

	0.9 mL	2 mL	300 µL	200 µL	50 µL	25 µL
H ₂ O weight in g.....					
1.						
2.						
3.						
Mean						
SD						

ID/ Date:

Kit Receipt

EpiDerm kit received (day/date/hour):	Day used:
EpiDerm (EPI-200-SCT) Lot no.: No. of inserts:	Production date:
Assay medium (EPI-100-ASY) Lot no.:	Expiration date:
MTT concentrate (MTT-100-CON); 2 ml Lot no.:	Expiration date:
MTT diluent (MTT-100-DIL); 8 ml Lot no.:	Production date:
MTT extractant (MTT-100-EXT), 60 ml Lot no.:	Expiration date:
PBS (TC-PBS); 100 ml Lot no.:	Expiration date:
No. of 6 well plates:	
No. of 24 well plates	
MTT protocol + packaging diagram	
Position of Ice-packs: (direct contact of the ice-packs with the skin must be avoided)	
Other remarks	

Date:.....ID

Quality Control of the Skin

Use scores: 1- very good, 2-good, 3- acceptable, 4- not acceptable

APPERANCE	KIT 1	KIT 2	KIT 3	KIT 4
MACRO.				
MICRO				
OTHER as. -edge defects -air bubbles -wet tissues --				

If you have any special observations (listed in OTHER), make reference to related tissue(s)

Kit No.1:.....

1	2	3	4	5	6
7	8	9	10	11	12
13	14	15	16	17	18
19	20	21	22	23	24

Kit No.2:.....

1	2	3	4	5	6
7	8	9	10	11	12
13	14	15	16	17	18
19	20	21	22	23	24

Kit No.3:.....

1	2	3	4	5	6
7	8	9	10	11	12
13	14	15	16	17	18
19	20	21	22	23	24

Kit No.4:.....

1	2	3	4	5	6
7	8	9	10	11	12
13	14	15	16	17	18
19	20	21	22	23	24

ID/ Date:

MTT Assay

Remarks before and after incubation with MTT

Before MTT incubation

3 min exposure		1 hour exposure	
Chemical tested	Observation (tissue #)	Chemical tested	Observation (tissue #)

After MTT incubation

3 min exposure		1 hour exposure	
Chemical tested	Observation (tissue #)	Chemical tested	Observation (tissue #)

*record observation as separation of the tissue from the insert during the washing procedure, reaction between insert and tested chemical, MTT reduction by chemical itself etc.

Date:.....ID.....

Measurement

Check plate photometer filter

Tick correct (✓) filter setting

reading filter: 570 nm	
no reference filter	

Date:.....ID.....

Archivation

Raw data saved in/as:

.....

Spreadsheet saved in/as:

.....

MDS saved in/as:

.....

Date:.....ID.....

ANNEX B: Test for Interference of Chemicals with MTT Endpoint and Correction Procedures

Possible interactions between test chemicals and test system

	Water coloration (Step 1, Section 7.3)	Tissue Staining (Step 2, Section 7.3)	MTT Interaction (Step 3, Section 7.3)	Test Conditions
Case 1	-	-	-	A
Case 2	+	-	-	A
Case 3	-	+	-	B
Case 4	+	+	-	B
Case 5	-	-	+	C
Case 6	+	-	+	C
Case 7	-	+	+	B+C
Case 8	+	+	+	B+C

Test Conditions:

A – Perform all steps according to the basic SOP. Correction of results using additional controls is not needed,
 B – Perform Step 2, Section 7.3 in addition to the basic SOP. Correction of results with viable tissue is needed.
 C – Perform Step 4, Section 7.3 in addition to the basic SOP. Correction of results with frozen tissue is needed.

B+C:

A colored chemical (or a chemical that may turn colored after interaction with water) may cause both tissue staining and false MTT reduction. If the experimentator is interested only in the correction of final results, combination of test condition C is sufficient for this purpose, since the frozen tissues absorb approximately the same amount of chemicals (and color) as the viable tissue. However, since the frozen tissues are more hydrated than viable tissues, tissue staining by water soluble colorants (or non-colored chemicals that will turn in aqueous conditions into colored) can be overestimated. In addition, conditions may arise when information on the amount of non-specific coloration is required. For increased precision of the above mentioned case, Steps 2 & 4 (Section 7.3) need to be performed.